Subsurface Wadden Sea sediments harbour an active C A R L V O N O S S I E T Z K Y universität and diverse community of sulfate-reducing bacteria OLDENBURG



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- Introduction



Fig. 1 Sampling site in the backbarrier of the island Spiekeroog,

Southern North Sea, Germany

Recent studies on intertidal flats give first insights into the structure and distribution of microbial communities in sediments up to several meters depth (Köpke et al. 2005, Wilms et al. 2006).

Sulfate-reducing bacteria (SRB) were detected by both cultivation and molecular methods. But still little is known about their activity and abundance in these deep marine sediments. Additionally, only few SRB from these depths were brought into pure culture so far. Here, we present geochemical and activity analyses combined with the selective cultivation of abundant SRB and their in situ



Results

quantification.



In situ abundance of cultured SRB

16 S rRNA-based DGGE and sequence analysis





Fig. 2 (A) Microbial sulfate reduction rates; (B) Pore water concentrations of sulfate; (C) Pore water concentrations of acetate and lactate as potential substrates for microbial sulfate reduction



Fig. 3 (A) Total DAPI Counts and absolut numbers of SRB as detected with CARD-FISH; (B) Relative abundance of different phylogenetic groups of SRB

Activity of SRB was highest in surface sediments, but was still detectable in several meters depth (SRR, Fig. 2A) In some layers, rates of microbial sulfate

reduction were close to those measured in the top-layer

Cultivation of abundant SRB

Significant sulfide production and, therefore, phylogenetic Sequencing and analysis of DGGE bands received from growth of SRB was observed in cultures from the highest sulfide-positive dilutions all depths and with lactate, acetate, or hydrogen als electron donor up to dilutions of revealed 13 partial 16S rRNA genes, most of them affiliating with marine 1:106 SRB of the *Deltaproteobacteria* (Fig. 4)

> uncultured delta proteobacterium Eel-BE1C3, AF354147 uncultured bacterium gas hydrate. AY053490 uncultured delta proteobacterium, gas hydrate, AJ535247 unc delta proteobacterium Eel-BE1B3, AF354151 uncultured delta proteobacterium, ZnS-prducing biofilm, AY082457 cultured delta proteobacterium Eel-BE1B3, AF354163 uncultured delta proteobacterium, Haakon Mosby sediment, AJ704678

Concentrations of potential substrates for microbial sulfate reduction were generally low (acetate: $< 15 \,\mu$ M) or, for lactate, showed local minima at several depths (Fig. 2C)

Both total cell counts and SRB counts varied only slightly with respect to the sediment depth (Fig. 3A)

SRB were found in all sediment layers accounting for up to 7 % of total cells

The most abundant SRB were members Desulfobacteraceae of the and the Desulfobulbaceae that accounted for at least two-thirds of all detected SRB in deep sediment layers (Fig. 3B)

Isolation of SRB into pure cultures

So far, 10 SRB originating from different sediment depths were isolated into pure cultures with lactate, acetate, or hydrogen as electron donor

Their phylogeny and physiology is presently under investigation



Fig. 5 Micrographs of representative morphotypes from pure cultures (A) 100 cm, hydrogen; (B) 250 cm, acetate, (C) 400 cm, hydrogen



Fig. 4 Phylogenetic affiliation of extracted DGGE bands (~500bp) received from the highest dilutions showing sulfide production. Sequences from this study are highlighted. The scale bar corresponds to 10% estimated sequence divergence.



Activity of sulfate-reducing bacteria in deep Wadden Sea sediments was demonstrated via radiotracer measurements

+ It was shown that active sulfate-reducing bacteria form a major fraction of the microbial community particularly in deep sediment layers

We successfully cultivated sulfate-reducing bacteria affiliating with as yet uncultured organisms exclusively detected by molecular approaches

+ Frequent cultivation of members of the *Desulfobacteraceae* and *Desulfobulbaceae* in dilution cultures was mirrored by their abundance in situ

Acknowledgements We wish to thank H. Nicolai, B. Engelen and F. Mathes for help in recovering sediment cores. Special thanks are given to S. Reischke for assistance in CARD-FISH and DGGE analyses. This work was financially supported by the Deutsche Forschungsgesellschaft (DFG).

Köpke et al. (2005) AEM 71:7819-7830 References Wilms et al. (2006) Environ Microbiol 8: 709-719